Clinical and Experimental Immunology REVIEW ARTICLE

doi:10.1111/j.1365-2249.2009.03944.x

Interleukin-17 and systemic lupus erythematosus: current concepts

A. Nalbandian, J. C. Crispín and G. C. Tsokos

Department of Medicine, Division of Rheumatology, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA, USA

Accepted for publication 24 March 2009 Correspondence: G. C. Tsokos, Professor of Medicine, Harvard Medical School, Chief, Rheumatology Division, Beth Israel Deaconess Medical Center, 330 Brookline Avenue, CLS-937, Boston, MA 02215, USA. E-mail: gtsokos@bidmc.harvard.edu

Summary

The emerging role of interleukin (IL)-17 as a hallmark proinflammatory cytokine of the adaptive immune system, produced primarily by a new T helper cell subset termed 'Th17', has received considerable attention. Differentiation of Th17 cells is driven by the simultaneous presence of transforming growth factor-β and certain inflammatory cytokines (e.g. IL-6, IL-21), and recent studies have shown that inflammation instigated by IL-17-producing cells is central to the development and pathogenesis of several human autoimmune diseases and animal models of autoimmunity. In this review, we focus on the information regarding IL-17 and systemic lupus erythematosus (SLE), a chronic autoimmune disease. The work that has explored the development and behaviour of IL-17-producing cells in SLE is discussed, and different mechanisms by which IL-17 could potentially augment inflammation and autoantibody production in the context of SLE are proposed.

Keywords: autoimmunity, DN T cells, IL-17, SLE, Th17

Interleukin-17-producing T cells and autoimmunity

Originally termed cytotoxic T lymphocyte antigen 8, interleukin (IL)-17 is a 17 kDa type I transmembrane protein isolated initially from a rodent CD4 T cell cDNA library [1]. It represents the prototype of a recently identified family of cytokines that comprises six members (IL-17A, IL-17B, IL-17C, IL-17D, IL-17E, IL-17F) and five receptors (IL-17RA, IL-17RB, IL-17RC, IL-17RD, IL-17RE) [2-4]. The most closely related members of the family are IL-17A and IL-17F, both of which are produced mainly by activated T cells [5] and bind to the same receptors (IL-17RA and IL-17RC) [6]. Thus, the dissection of their individual contributions to normal and abnormal immune responses has been complex and is not understood completely [7,8]. Although the signalling pathway downstream of IL-17R remains to be elucidated, recent studies have shown that nuclear factor-κB and mitogen-activated protein kinase pathways are involved [9,10].

In addition to its potent proinflammatory capacity, IL-17 exerts its effects through the recruitment of monocytes and neutrophils by increasing the local production of chemokines (IL-8, monocyte chemoattractant protein-1, growth-related oncogene protein- α) [11–15], the facilitation of T cell infiltration and activation by stimulating the expression of intercellular adhesion molecule-1 [16] as well as the amplification of the immune response by inducing the production of IL-6, prostaglandin E2, granulocyte-macrophage colonystimulating factor and granulocyte colony-stimulating factor [17,18]. Additionally, IL-17 synergizes with other cytokines, in particular with IL-1 β , tumour necrosis factor (TNF)- α , and interferon (IFN)- γ [19–22]. IL-17RA is expressed broadly and mediates its effects through a number of immune and non-immune cells (particularly endothelial and epithelial

The IL-17 is produced by several cell types that include CD4⁺ T cells, CD8⁺ T cells, CD3⁺CD4⁻CD8⁻ T cells, γδ-T cells, natural killer cells and neutrophils [24]. IL-17 plays an important role in the immune response against certain pathogens, particularly bacteria (e.g. Klebsiella pneumoniae, Citrobacter rodentium, Borrelia burgdorferi) [25-27] and fungi (e.g. Candida albicans) [28]. In concordance, human and murine antigen-presenting cells (APC) induce the differentiation of naive T cells into IL-17-producing cells when incubated with certain microbial products such as lipopolysaccharide, peptidoglycans and zymosan [29,30].

The CD4+ T cell effector subset termed 'Th17' has been considered a remarkable discovery which was named after its signature cytokine, IL-17 [31]. Th17 cells are considered as a distinct T helper cell subset because: (i) they arise from naive T cells when primed in the presence of specific factors; (ii) their differentiation is controlled by exclusive transcription factors; (iii) they exhibit a particular cytokine production

profile; and (iv) their differentiation into Th17 cells excludes the acquisition of other effector phenotypes (i.e. Th1 and Th2). The concept of Th17 cells was conceived within the setting of experimental autoimmune diseases and these cells have, since their first description, been associated intimately with autoimmune responses.

The IL-23 is a heterodimer consisted of two subunits, p19 and p40 [32]. Although p19 is exclusive to IL-23, p40 is also part of IL-12 (when combined with p35). The discovery of p19, in 2000, implied that p40-deficient mice were not only IL-12-deficient (as considered hitherto) but also IL-12and IL-23-deficient. The subsequent comparison between IL-23p19-deficient mice and IL-12p35-deficient mice demonstrated that the absence of IL-23 (rather than the absence of IL-12 and thus Th1 cells) protected mice from experimental autoimmune encephalitis (EAE) [33]. IL-23 was necessary for the development of a pathogenic Th17 response that could be transferred into naive wild-type mice upon T cell adoptive transfer [34]. These discoveries, along with the fact that IL-23 increases IL-17 production by memory T cells [23], suggested the presence of a novel T helper functional axis formed by IL-23 (as an IL-17-inducing APC-derived factor) and IL-17 (as the product of the differentiated T cell) analogous to the IL-12/Th1 system. Further work demonstrated that naive T cells lack IL-23R and that this cytokine contributes to the expansion and maintenance of the Th17 subset rather than to its differentiation [31,35].

The combination of IL-6 and transforming growth factor (TGF)-β was shown to induce the differentiation of murine naive T cells into Th17 cells [36-38]. This notion is extremely interesting, as it implies that the presence of an inflammatory signal (i.e. IL-6) is the key element that determines whether naive T cells become proinflammatory (i.e. Th17) or suppressive (regulatory T cells). Accordingly, IL-6deficient mice are resistant to EAE induction and have defective Th17 cell differentiation [37,39]. The specific contribution of TGF-β, which is apparently able to induce both suppressive and inflammatory cells, as well as the particular instances when it participates in T cell priming, will need to be addressed in future studies. IL-21, a cytokine related to IL-2 [40], is also able to induce T cell differentiation into Th17 cells [41-43]. Contrary to IL-6, IL-21 is not produced by APCs but by T cells - particularly Th17 cells and T follicular helper cells (T_{FH}) – and thus has been postulated to act as an autoamplifier of the Th17 response [41-43]. Differentiation of human naive T cells into Th17 cells is accomplished similarly when TGF- β is present along with IL-21, or different combinations of IL-6, IL-23, IL-1 β and TNF- α [44–46].

The genetic programme responsible for driving the Th17 phenotype is accomplished by at least three transcription factors. Two of them belong to the family of retinoid-related orphan receptors (RORγt and RORα). RORγt is expressed selectively in Th17 cells and its presence is necessary for IL-17A and IL-17F production [47]. RORα, however, can also promote Th17 differentiation [48]. Both factors are induced by TGF-β and IL-6 through a signal transducer and activator transcription-3-dependent mechanism [48]. The relative contribution of these transcription factors to Th17 differentiation and behaviour, during normal and abnormal immune responses, is still unknown.

In summary, IL-17 is a potent proinflammatory cytokine produced by activated T cells, particularly Th17 cells. Although necessary in the responses against bacteria and fungi, IL-17 has been associated with the pathogenesis of a wide range of inflammatory and autoimmune diseases including psoriasis, rheumatoid arthritis (RA) [49,50], inflammatory bowel disease [51], systemic sclerosis [52] and systemic lupus erythematosus (SLE) [53,54].

Murine models of IL-17 and SLE

From a conceptual point of view, IL-17 and Th17 cells were regarded initially as potentially involved in diseases considered formerly to be driven by Th1 cells. Thus, early investigations focused upon EAE [34] and murine models of RA [55,56], and it was found that inhibition of IL-17 (or IL-17producing cells) was beneficial. Th17 cells were eagerly considered part of the pathogenesis of such diseases as powerful effector cells able to amplify organ-specific destructive inflammatory responses by means of producing chemokines and cytokines. Although SLE has been considered classically an autoantibody- and immune complex-driven disease, recent work indicates that IL-17 is involved in different aspects of its pathogenesis. Its potent proinflammatory capacity, along with the effects it exerts in a variety of cells, suggests that its unregulated production has indeed widespread consequences in animals and patients with lupus (Tables 1 and 2). Lupus-prone mice (MRL/lpr) are particularly susceptible to the development of inflammation induced by ischaemic insults [57]. We have shown that

Table 1. Interleukin (IL)-17 in murine models of systemic lupus ervthematosus.

Murine models	Experimental evidence and outcomes	References
MRL/lpr	Enhanced IL-17-mediated tissue injury after ischaemia reperfusion	Edgerton et al., 2009 [58]
BXD2	Increased numbers of IL-17-producing T cells provide help to B cells and stimulate spleen germinal centre formation. IL-17 over-expression enhanced disease; IL-17R blockade reduced its intensity	Hsu et al., 2008 [62]
SNF_1	Increased numbers of IL-17 ⁺ cells	Kang et al., 2007 [59]
Ets-1 knock-out	Enhanced differentiation of naive T cells into Th17 cells	Wang et al., 2005 [68]

Table 2. Interleukin (IL)-17 in human SLE.

Experimental evidence and outcomes	References	
Significant levels of IL-17 and IFN-γ detected in double-negative T cells from SLE patients	Crispin et al., 2008 [53]	
Elevated levels of IL-23 and IL-17 in sera from SLE patients	Wong et al., 2008 [54]	
IL-17 increases autoantibody production from PBMC in patients from lupus nephritis	Dong et al., 2003 [64]	

IFN, interferon; SLE, systemic lupus erythematosus; PBMC, peripheral blood mononuclear cells.

intestinal injury following ischaemia and reperfusion is mediated, at least partially, by CD4⁺ T cells that produce IL-17, and that this phenomenon is augmented in MRL/lpr mice [58]. Accordingly, CD4⁺ T cell depletion suppressed injury induced by ischaemia and deficiency of IL-17 (in IL-23p19^{-/-} mice) reduced tissue injury significantly. The reduction in tissue injury observed in the absence of IL-17 was more pronounced in MRL/lpr mice than in non-autoimmune B6 mice [58]. This effect is probably due to the fact that MRL/lpr mice have a higher frequency of IL-17-producing T cells (unpublished data, Tsokos Laboratory).

Splenocytes from lupus-prone SNF₁ mice (New Zealand black × SWR F₁) produce significantly higher amounts of IL-17 than splenocytes from non-autoimmune C57Bl/6J (B6) mice when cultured in the presence of nucleosomes [59]. Moreover, IL-17-producing T cells were detected in kidneys affected by nephritis in SNF₁ mice. Importantly, clinical improvement achieved either by tolerance induction with a histone-derived peptide [59] or with nasal administration of anti-CD3 [60] was accompanied by decreased IL-17 production, decreased percentage of IL-17-producing cells and abrogation of IL-17+ kidney-infiltrating cells [59]. Interestingly, in the first of these reports, tolerance induction was associated with decreased IL-6 production and increased TGF- β production that paralleled a reduction in the fraction of IL-17-producing T cells and a reciprocal increase in regulatory T cells [59]. This highlights the fact that the SLE microenvironment is particularly apt for the differentiation of IL-17-producing cells.

The BXD2 mouse, a lupus model that develops arthritis, glomerulonephritis and autoantibodies spontaneously [61], has high IL-17 levels in serum as well as increased numbers of IL-17⁺ cells in the spleen [62]. Accordingly, upon stimulation an increased fraction of BXD2 T cells produce IL-17. The humoral response is augmented strongly in these mice [63]. They develop germinal centres (GC) spontaneously in the spleen, where IL-17⁺ T cells co-localize with IL-17R⁺ B cells [62]. The importance of IL-17 in this process was demonstrated when B6 and pre-disease BXD2 mice were infected with an IL-17-coding adenovirus that increased IL-17 levels and induced the formation of GC in both mouse strains. Concordantly, formation of GC diminished and production of anti-DNA and anti-histone antibodies was abrogated in BXD2 IL-17R-deficient mice [62]. This study indicates that besides acting as a mediator of inflammation, IL-17 can also provide help to B cells, a notion suggested by a former report that showed that IL-17 increased immunoglobulin (Ig)G

and anti-DNA antibody production in mononuclear cells derived from SLE patients [64].

Intriguingly, a recent paper describes a T cell effector population akin to T_{FH} [65] that provides help to B cells in an extrafollicular location, a phenomenon observed in several lupus-prone strains (e.g. MRL/lpr and NZB \times W F₁) [66]. Extrafollicular helper cells provide B cell help via IL-21 and CD40L and their development depends upon the presence of inducible co-stimulatory molecule [66]. Along with IL-21, these cells produce IL-17, but IL-17 does not seem to play an essential role in B cell stimulation [66]. On the other hand, the IL-17-inducing capacity of IL-21 has been proposed to contribute to the autoimmune response, because mice with deficient IL-21 signalling have reduced numbers of Th17 cells [41]. However, in BXSB-Yaa mice, IL-21R deficiency abrogated autoimmune disease without affecting IL-17 levels or reducing the frequency of IL-17-producing cells [67]. Thus, although IL-21 derived from follicular helper T cells seems to play a role in B cell stimulation in autoimmune murine models, its contribution to Th17 cell differentiation is still debatable. Similarly, although in certain conditions IL-17 has proved to be able to provide B cell help, the precise contribution of IL-17 derived from T_{FH} cells to GC formation and antibody production will have to be defined more clearly.

The Ets-1 knock-out mouse is another mouse model which demonstrates a potential pathogenic role for IL-17-producing T cells. Ets-1^{-/-} mice display high levels of IgG and IgM autoantibodies, leading further to the deposition of immune complexes in kidney glomeruli and complement activation [68]. Although elevated serum levels were not detected in these Ets-1 knock-out mice, increased levels of IL-17A, IL-17F and IL-22 mRNA were found in the lung, consistent with process of inflammation in this tissue.

A recent report grants TNF- α a protective role in SLE. Jacob *et al.* studied NZM2328 mice deficient in both TNF- α receptors and found that disease severity (in terms of nephritis and production of anti-DNA antibodies) increased when the TNF- α pathway was disrupted. The effect depended upon the presence of CD4⁺ T cells that exhibited a Th17 gene profile. This suggests that acceleration of nephritis in SLE may indeed be associated with the IL17/Th17 pathway [69].

Mechanisms of autoimmune pathology: IL-17 and human SLE

The SLE is a complex autoimmune disease in which a T cell-driven autoimmune response against ubiquitously

expressed autoantigens results in clinically and pathologically diverse manifestations [70]. Although the presence of a large array of autoantibodies is perhaps the most conspicuous characteristic of SLE patients, target organ infiltration and chronic inflammation are essential pathogenic features that result in end-organ damage in most SLE clinical manifestations (i.e. nephritis, vasculitis, discoid lupus) [71]. Recent evidence indicates that IL-17 plays a role in the pathogenesis of SLE [72]. SLE patients have higher serum levels of IL-17 and IL-23 than healthy controls [54,73]. Moreover, the frequency of IL-17-producing T cells is increased in peripheral blood of SLE patients [53,54]. Accordingly, IL-17 production is increased in in vitro stimulated lymphocytes from SLE patients when compared with normal lymphocytes [54]. Plasma IL-17 levels show a positive correlation with SLE disease activity [54].

In a recent work, we demonstrated that a significant fraction of the IL-17 produced in SLE patients derives from double-negative (DN) TCR- $\alpha\beta^+$ CD4 $^-$ CD8 $^-$ T cells [53]. Scarce in healthy individuals, DN T cells are expanded in peripheral blood of SLE patients and produce IL-17 and IFN- γ . Furthermore, DN T cells and IL-17-producing T cells are found in kidney biopsies from patients with lupus nephritis [53]. Along with IL-13, IFN- γ and IL-17 were the main cytokines produced by infiltrating T cells in nephritic kidneys of MRL/lpr mice [74]. The finding of DN T cells within a T cell infiltrate demonstrates their capacity to accumulate in inflamed tissue and suggests strongly that they play a pathogenic role in the local inflammatory response [53]. Similarly, the demonstration of IL-17 $^+$ T cells in kidneys affected by lupus nephritis indicates that it may play a role in

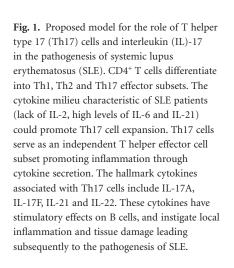
the amplification and perpetuation of the inflammatory response in organs targeted by SLE.

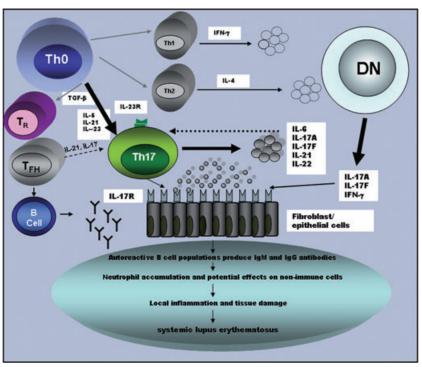
Apart from the obvious proinflammatory activities attributed to IL-17, its effects in other cell types may contribute to SLE pathogenesis. Accordingly, increased production of total IgG, anti-dsDNA IgG and IL-6 by peripheral blood mononuclear cells of patients with lupus nephritis was observed when they were cultured with IL-17 [64]. These findings suggest that IL-17 may participate in the activation of B cells in patients with SLE (Table 2).

Concluding remarks and perspectives

In this communication, we have reviewed reports that address the relationship between IL-17 and SLE. Virtually, all papers indicate that IL-17 production is increased in patients with SLE as well as in animals with lupus-like diseases. This could be a consequence of systemic inflammation and augmented T cell activation [75], or could indicate that the pathways that guide T cell differentiation into IL-17-producing cells (either Th17 or DN T cells) are facilitated in SLE patients (Fig. 1). This possibility is plausible because IL-6 [76] and IL-21 production (unpublished data, Tsokos Laboratory) has been found increased in patients with SLE. Moreover, the reciprocal regulatory T cell deficiency reported in SLE patients [77] could also be a consequence of skewed T cell differentiation. These issues will have to be addressed specifically in future studies.

Evidence obtained in human samples places IL-17 in the midst of the inflammatory reaction in SLE patients [53]. Although a causal association cannot be sought in human





studies, animal models have demonstrated that blockade of IL-17 decreases lupus manifestations [62]. Interestingly, information obtained in these studies suggests that IL-17 could be associated not only with T cell-mediated tissue injury but also with production of pathogenic autoantibodies. SLE-derived B cells have been shown to increase anti-DNA production when cultured in the presence of IL-17 [64]. The relative importance of this pathogenic mechanism in human SLE remains to be demonstrated in future work.

The evidence provided in this review describes IL-17 as an important cytokine in the pathogenesis of SLE. Its exact place within the mechanisms that lead to SLE remains to be defined. IL-17 might represent an effector cytokine associated to tissue damage and disease amplification or perhaps a cytokine whose abnormal presence during otherwise normal immune responses causes tolerance disruption. These issues, as well as the main question of whether IL-17 blockade will be therapeutically useful for SLE patients, will be addressed in the near future [78,79].

Acknowledgements

Work performed in the authors' laboratory was supported by National Institutes of Health Grants R01 AI043043 and T32 AI074549 as well as by the Mary Kirkland Center for Lupus Research at the Hospital for Special Surgery funded by Rheuminations.

References

- 1 Rouvier E, Luciani MF, Mattei MG, Denizot F, Golstein P. CTLA-8, cloned from an activated T cell, bearing AU-rich messenger RNA instability sequences, and homologous to a herpesvirus saimiri gene. J Immunol 1993; 150:5445–56.
- 2 Aggarwal S, Gurney AL. IL-17: prototype member of an emerging cytokine family. J Leuk Biol 2002; 71:1–8.
- 3 Yao Z, Timour M, Painter S, Fanslow W, Spriggs M. Complete nucleotide sequence of the mouse CTLA8 gene. Gene 1996; 168:223–5
- 4 Weaver CT, Hatton RD, Mangan PR, Harrington LE. IL-17 family cytokines and the expanding diversity of effector T cell lineages. Annu Rev Immunol 2007; 25:821–52.
- 5 Chen Z, O'Shea JJ. Regulation of IL-17 production in human lymphocytes. Cytokine 2008; **41**:71–8.
- 6 Toy D, Kugler D, Wolfson M et al. Cutting edge: interleukin 17 signals through a heteromeric receptor complex. J Immunol 2006; 177:36–9.
- 7 Kolls JK, Linden A. Interleukin-17 family members and inflammation. Immunity 2004; **21**:467–76.
- 8 Dubin PJ, Kolls JK. Interleukin-17A and interleukin-17F: a tale of two cytokines. Immunity 2009; **30**:9–11.
- 9 Schwandner R, Yamaguchi K, Cao Z. Requirement of tumor necrosis factor receptor-associated factor (TRAF)6 in interleukin 17 signal transduction. J Exp Med 2000; 191:1233–40.
- 10 Shalom-Barak T, Quach J, Lotz M. Interleukin-17-induced gene expression in articular chondrocytes is associated with activation of mitogen-activated protein kinases and NF-kappaB. J Biol Chem 1998; 273:27467–73.

- 11 Laan M, Lotvall J, Chung KF, Linden A. IL-17-induced cytokine release in human bronchial epithelial cells in vitro: role of mitogen activated protein (MAP) kinases. Br J Pharmacol 2001; 133:200–6.
- 12 Woltman AM, de Haij S, Boonstra JG, Gobin SJ, Daha MR, van Kooten C. Interleukin-17 and CD40-ligand synergistically enhance cytokine and chemokine production by renal epithelial cells. J Am Soc Nephrol 2000; 11:2044–55.
- 13 Witowski J, Pawlaczyk K, Breborowicz A et al. IL-17 stimulates intraperitoneal neutrophil infiltration through the release of GRO alpha chemokine from mesothelial cells. J Immunol 2000; 165:5814–21.
- 14 Ruddy MJ, Shen F, Smith JB, Sharma A, Gaffen SL. Interleukin-17 regulates expression of the CXC chemokine LIX/CXCL5 in osteo-blasts: implications for inflammation and neutrophil recruitment. J Leukoc Biol 2004; 76:135–44.
- 15 Agarwal S, Misra R, Aggarwal A. Interleukin 17 levels are increased in juvenile idiopathic arthritis synovial fluid and induce synovial fibroblasts to produce proinflammatory cytokines and matrix metalloproteinases. J Rheumatol 2008; 35:515–9.
- 16 Albanesi C, Cavani A, Girolomoni G. IL-17 is produced by nickel-specific T lymphocytes and regulates ICAM-1 expression and chemokine production in human keratinocytes: synergistic or antagonist effects with IFN-gamma and TNF-alpha. J Immunol 1999; 162:494–502.
- 17 Schwarzenberger P, Huang W, Ye P et al. Requirement of endogenous stem cell factor and granulocyte-colony-stimulating factor for IL-17-mediated granulopoiesis. J Immunol 2000; 164:4783–9.
- 18 Cai X, Gommoll Jr CP, Justice L, Narula SK, Fine JS. Regulation of granulocyte colony-stimulating factor gene expression by interleukin-17. Immunol Lett 1998; 62:51–8.
- 19 Laan M, Cui ZH, Hoshino H et al. Neutrophil recruitment by human IL-17 via C-X-C chemokine release in the airways. J Immunol 1999; 162:2347–52.
- 20 Maertzdorf J, Osterhaus AD, Verjans GM. IL-17 expression in human herpetic stromal keratitis: modulatory effects on chemokine production by corneal fibroblasts. J Immunol 2002; 169:5897–903.
- 21 Van Bezooijen RL, Papapoulos SE, Lowik CW. Effect of interleukin-17 on nitric oxide production and osteoclastic bone resorption: is there dependency on nuclear factor-kappaB and receptor activator of nuclear factor kappaB (RANK)/RANK ligand signaling? Bone 2001; 28:378–86.
- 22 Ruddy MJ, Wong GC, Liu XK et al. Functional cooperation between interleukin-17 and tumor necrosis factor-alpha is mediated by CCAAT/enhancer-binding protein family members. J Biol Chem 2004; 279:2559–67.
- 23 Aggarwal S, Ghilardi N, Xie MH, de Sauvage FJ, Gurney AL. Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. J Biol Chem 2003; 278:1910–4.
- 24 Korn T, Oukka M, Kuchroo VK, Bettelli E. Th17 cells: effector cells with inflammatory properties. Semin Immunol 2007; 19:362–71.
- 25 Happel KI, Dubin PJ, Zheng M et al. Divergent roles of IL-23 and IL-12 in host defense against Klebsiella pneumoniae. J Exp Med 2005; 202:761–9.
- 26 Zheng Y, Valdez PA, Danilenko DM et al. Interleukin-22 mediates early host defense against attaching and effacing bacterial pathogens. Nat Med 2008; 14:282–9.
- 27 Infante-Duarte C, Horton HF, Byrne MC, Kamradt T. Microbial lipopeptides induce the production of IL-17 in Th cells. J Immunol 2000; 165:6107–15.

- 28 Huang W, Na L, Fidel PL, Schwarzenberger P. Requirement of interleukin-17A for systemic anti-*Candida albicans* host defense in mice. J Infect Dis 2004; 190:624–31.
- 29 LeibundGut-Landmann S, Gross O, Robinson MJ *et al.* Syk- and CARD9-dependent coupling of innate immunity to the induction of helper T cells that produce interleukin 17. Nat Immunol 2007; **8**:630–8.
- 30 Acosta-Rodriguez EV, Rivino L, Geginat J et al. Surface phenotype and antigenic specificity of human interleukin 17-producing T helper memory cells. Nat Immunol 2007; 8:549–51.
- 31 Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 cells. Annu Rev Immunol 2009.
- 32 Oppmann B, Lesley R, Blom B *et al.* Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. Immun 2000; **13**:715–25
- 33 Cua DJ, Sherlock J, Chen Y et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. Nature 2003; 421:744–8.
- 34 Langrish CL, Chen Y, Blumenschein WM *et al.* IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. J Exp Med 2005; **201**:233–40.
- 35 Kastelein RA, Hunter CA, Cua DJ. Discovery and biology of IL-23 and IL-27: related but functionally distinct regulators of inflammation. Annu Rev Immunol 2007; 25:221–42.
- 36 Veldhoen M, Hocking RJ, Atkins CJ, Locksley RM, Stockinger B. TGFbeta in the context of an inflammatory cytokine milieu supports de novo differentiation of IL-17-producing T cells. Immunity 2006; 24:179–89.
- 37 Bettelli E, Carrier Y, Gao W *et al.* Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. Nature 2006; **441**:235–8.
- 38 Mangan PR, Harrington LE, O'Quinn DB *et al.* Transforming growth factor-beta induces development of the T(H)17 lineage. Nature 2006; **441**:231–4.
- 39 Samoilova EB, Horton JL, Hilliard B, Liu TS, Chen Y. IL-6-deficient mice are resistant to experimental autoimmune encephalomyelitis: roles of IL-6 in the activation and differentiation of autoreactive T cells. J Immunol 1998; 161:6480–6.
- 40 Parrish-Novak J, Dillon SR, Nelson A et al. Interleukin 21 and its receptor are involved in NK cell expansion and regulation of lymphocyte function. Nature 2000; 408:57–63.
- 41 Korn T, Bettelli E, Gao W et al. IL-21 initiates an alternative pathway to induce proinflammatory T(H)17 cells. Nature 2007; 448:484-7.
- 42 Nurieva R, Yang XO, Martinez G *et al.* Essential autocrine regulation by IL-21 in the generation of inflammatory T cells. Nature 2007; **448**:480–3.
- 43 Zhou L, Ivanov II, Spolski R *et al.* IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways. Nat Immunol 2007; **8**:967–74.
- 44 Yang L, Anderson DE, Baecher-Allan C *et al.* IL-21 and TGF-beta are required for differentiation of human T(H)17 cells. Nature 2008: **454**:350–2.
- 45 Manel N, Unutmaz D, Littman DR. The differentiation of human T(H)-17 cells requires transforming growth factor-beta and induction of the nuclear receptor RORgammat. Nat Immunol 2008; 9:641–9.
- 46 Volpe E, Servant N, Zollinger R et al. A critical function for transforming growth factor-beta, interleukin 23 and proinflammatory

- cytokines in driving and modulating human T(H)-17 responses. Nat Immunol 2008; 9:650–7.
- 47 Ivanov II, McKenzie BS, Zhou L et al. The orphan nuclear receptor RORgammat directs the differentiation program of proinflammatory IL-17+ T helper cells. Cell 2006; 126:1121–33.
- 48 Yang XO, Pappu BP, Nurieva R et al. T helper 17 lineage differentiation is programmed by orphan nuclear receptors ROR alpha and ROR gamma. Immun 2008; 28:29–39.
- 49 Miossec P. Interleukin-17 in rheumatoid arthritis: if T cells were to contribute to inflammation and destruction through synergy. Arthritis Rheum 2003; 48:594–601.
- 50 Van bezooijen RL, Farih-Sips HC, Papapoulos SE, Lowik CW. Interleukin-17: a new bone acting cytokine *in vitro*. J Bone Miner Res 1999; 14:1513–21.
- 51 Shih DQ, Targan SR, McGovern D. Recent advances in IBD pathogenesis: genetics and immunobiology. Curr Gastroenterol Rep 2008; 10:568–75.
- 52 Kurasawa K, Hirose K, Sano H et al. Increased interleukin-17 production in patients with systemic sclerosis. Arthritis Rheum 2000; 43:2455–63.
- 53 Crispin JC, Oukka M, Bayliss G *et al.* Expanded double negative T cells in patients with systemic lupus erythematosus produce IL-17 and infiltrate the kidneys. J Immunol 2008; **181**:8761–6.
- 54 Wong CK, Lit LC, Tam LS, Li EK, Wong PT, Lam CW. Hyper-production of IL-23 and IL-17 in patients with systemic lupus erythematosus: implications for Th17-mediated inflammation in auto-immunity. Clin Immunol 2008; 127:385–93.
- 55 Sato K, Suematsu A, Okamoto K et al. Th17 functions as an osteoclastogenic helper T cell subset that links T cell activation and bone destruction. J Exp Med 2006; 203:2673–82.
- 56 Nakae S, Nambu A, Sudo K, Iwakura Y. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. J Immunol 2003; 171:6173–7.
- 57 Fleming SD, Monestier M, Tsokos GC. Accelerated ischemia/ reperfusion-induced injury in autoimmunity-prone mice. J Immunol 2004; 173:4230–5.
- 58 Edgerton C, Crispin JC, Moratz CM *et al.* IL-17 producing CD4+ T cells mediate accelerated ischemia/reperfusion-induced injury in autoimmunity-prone mice. Clin Immunol 2009; **130**:313–21.
- 59 Kang HK, Liu M, Datta SK. Low-dose peptide tolerance therapy of lupus generates plasmacytoid dendritic cells that cause expansion of autoantigen-specific regulatory T cells and contraction of inflammatory Th17 cells. J Immunol 2007; **178**:7849–58.
- 60 Wu HY, Quintana FJ, Weiner HL. Nasal anti-CD3 antibody ameliorates lupus by inducing an IL-10-secreting CD4+. J Immunol 2008; 181:6038–50.
- 61 Hsu HC, Zhou T, Kim H et al. Production of a novel class of polyreactive pathogenic autoantibodies in BXD2 mice causes glomerulonephritis and arthritis. Arthritis Rheum 2006; 54:343–55.
- 62 Hsu HC, Yang P, Wang J et al. Interleukin 17-producing T helper cells and interleukin 17 orchestrate autoreactive germinal center development in autoimmune BXD2 mice. Nat Immunol 2008; 9:166–75.
- 63 Hsu HC, Wu Y, Yang P et al. Overexpression of activation-induced cytidine deaminase in B cells is associated with production of highly pathogenic autoantibodies. J Immunol 2007; 178:5357–65.
- 64 Dong G, Ye R, Shi W et al. IL-17 induces autoantibody overproduction and peripheral blood mononuclear cell overexpression of IL-6 in lupus nephritis patients. Chin Med J (Engl) 2003; 116:543–8.

- 65 Breitfeld D, Ohl L, Kremmer E et al. Follicular B helper T cells express CXC chemokine receptor 5, localize to B cell follicles, and support immunoglobulin production. J Exp Med 2000; 192:1545– 52.
- 66 Odegard JM, Marks BR, DiPlacido LD et al. S-dependent extrafollicular helper T cells elicit IgG production via IL-21 in systemic autoimmunity. J Exp Med 2008; 205:2873–86.
- 67 Bubier JA, Sproule TJ, Foreman O et al. A critical role for IL-21 receptor signaling in the pathogenesis of systemic lupus erythematosus in BXSB-Yaa mice. Proc Natl Acad Sci USA 2009.
- 68 Wang D, John SA, Clements JL, Percy DH, Barton KP, Garrett-Sinha LA. Ets-1 deficiency leads to altered B cell differentiation, hyperresponsiveness to TLR9 and autoimmune disease. Int Immunol 2005; 17:1179–91.
- 69 Jacob N, Yang H, Pricop L et al. Accelerated pathological and clinical nephritis in systemic lupus erythematosus-prone New Zealand Mixed 2328 mice doubly deficient in TNF receptor 1 and TNF receptor 2 via a Th17-associated pathway. J Immunol 2009; 182:2532–41.
- 70 Crispin JC, Kyttaris VC, Juang YT, Tsokos GC. How signaling and gene transcription aberrations dictate the systemic lupus erythematosus T cell phenotype. Trends Immunol 2008; 29:110–5.
- 71 Cohen RA, Bayliss G, Crispin JC *et al.* T cells and *in situ* cryoglobulin deposition in the pathogenesis of lupus nephritis. Clin Immunol 2008; **128**:1–7.

- 72 Garrett-Sinha LA, John S, Gaffen SL. IL-17 and the Th17 lineage in systemic lupus erythematosus. Curr Opin Rheumatol 2008; 20:519–25.
- 73 Wong CK, Ho CY, Li EK, Lam CW. Elevation of proinflammatory cytokine (IL-18, IL-17, IL-12) and Th2 cytokine (IL-4) concentrations in patients with systemic lupus erythematosus. Lupus 2000; 9:589–93.
- 74 Wang Y, Ito S, Chino Y et al. Use of laser microdissection in the analysis of renal-infiltrating T cells in MRL/lpr mice. Mod Rheumatol 2008; 18:385–93.
- 75 Crispin JC, Martinez A, de PP, Velasquillo C, Alcocer-Varela J. Participation of the CD69 antigen in the T-cell activation process of patients with systemic lupus erythematosus. Scand J Immunol 1998: 48:196–200.
- 76 Decker P, Kotter I, Klein R, Berner B, Rammensee HG. Monocytederived dendritic cells over-express CD86 in patients with systemic lupus erythematosus. Rheumatology (Oxf) 2006; 45:1087–95.
- 77 Crispin JC, Martinez A, Alcocer-Varela J. Quantification of regulatory T cells in patients with systemic lupus erythematosus. J Autoimmun 2003; 21:273–6.
- 78 Ghilardi N, Ouyang W. Targeting the development and effector functions of TH17 cells. Semin Immunol 2007; 19:383–93.
- 79 Kikly K, Liu L, Na S, Sedgwick JD. The IL-23/Th(17) axis: therapeutic targets for autoimmune inflammation. Curr Opin Immunol 2006; 18:670–5.